

ALLTogether Consortium (ATG) protocol

CYTOGENETIC STRATIFICATION

UPDATES GUIDELINES octobre 2022

GFCH 1 février 2023

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GEN-GR
CYTO-GR

or CNA group A
w/o CYTO-HR

Good risk genetic abnormalities

Good risk cytogenetic abnormalities

- *ETV6-RUNX1*/t(12;21)(p13;q22)
- High Hyperdiploidy (51-65 chromosomes)

Good risk copy number alteration profiles

- No deletion of *IKZF1*, *CDKN2A/B*, *PAR1*, *BTG1*, *EBF1*, *PAX5*, *ETV6* or *RB1*
- Isolated deletions of *ETV6*, *PAX5* or *BTG1*
- *ETV6* deletions with a single additional deletion of *BTG1*, *PAX5* or *CDKN2A/B*

GEN-PR
CYTO-HR

or CNA group B
w/o CYTO-GR

Poor risk genetic abnormalities

High risk cytogenetic subgroups

- t(9;22)(q34;q11)/*BCR-ABL1*
- *MLL*/11q23 translocation
- Near haploidy (<30 chromosomes)
- Low hypodiploidy / near triploidy (30-39 / 60-78 chromosomes)
- Intrachromosomal amplification of chromosome 21 (iAMP21)
- t(17;19)(q23;p13)/*TCF3-HLF*

+ *Ig::MYC* Update Guidelines

Intermediate and poor risk copy number alteration profiles

- Any deletion of *IKZF1*, *PAR1*, *EBF1* or *RB1*
- All other copy number alteration profiles not mentioned above.

Inf ou égal à 16ans



Patients are classified hierarchically with cytogenetic abnormalities taking precedence over copy number alteration profiles.

Definition of novel genetic risk groups for pediatric BCP-ALL.

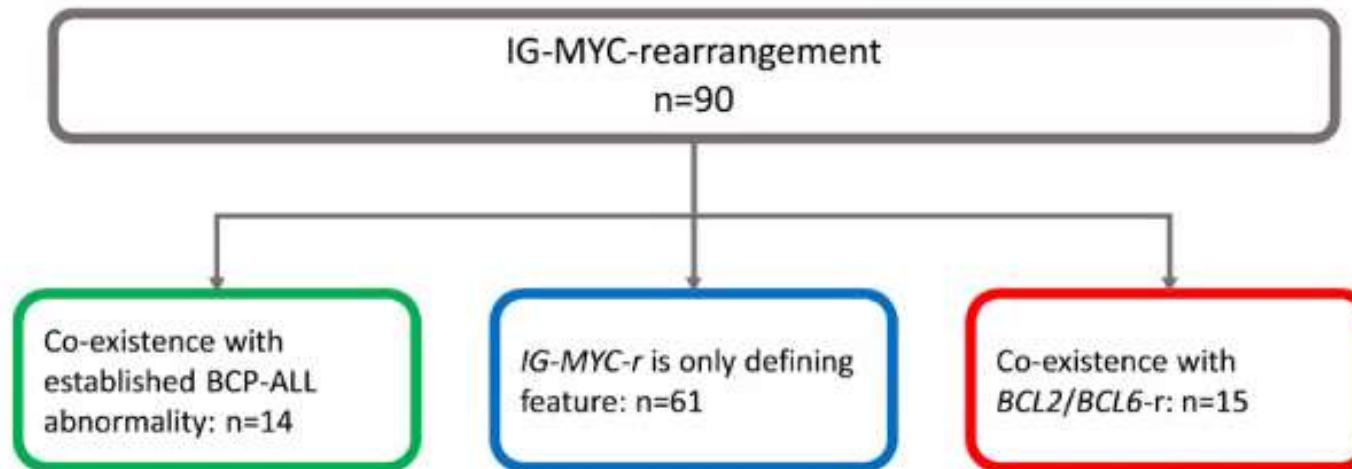
Molecular characterisation and clinical outcome of B-cell precursor ALL with Ig::MYC rearrangement

Bomken, et al, Haematologica 2022

- Approximately 0.5% patients who present with surface immunoglobulin negative BCP-ALL are characterised by a translocation between an immunoglobulin (IG) locus and MYC (being any of t(8;14)(q24;q32), t(2;8)(p12;q24) or t(8;22)(q24;q11))
- Defined as **having IG::MYC in the absence of other subtype** defining genetic abnormalities and without translocation of BCL2 or BCL6, this group has a poor event free survival.
- Patients with BCP-ALL and IG::MYC are **eligible for inclusion on the trial unless a concomitant BCL2/6 translocation is present.**
- **Some IG::MYC rearrangements are sub-clonal to other subtype defining genetic events** (e.g. KMT2A-r, high hyperdiploidy) → In this situation, stratification will be based on the primary genetic abnormality.

Molecular characterisation and clinical outcome of B-ALL with IG::MYC rearrangement

Bomken et al, Haematologica, 2022



*NB : IG::MYC may co-occur as a **sub clonal abnormality** to HeH, KMT2A-r, iAMP21, BCR:: ABL1, IGH::DUX4*

NB : Si 8q24 anormal au caryotype du diagnostic toujours penser à MYC

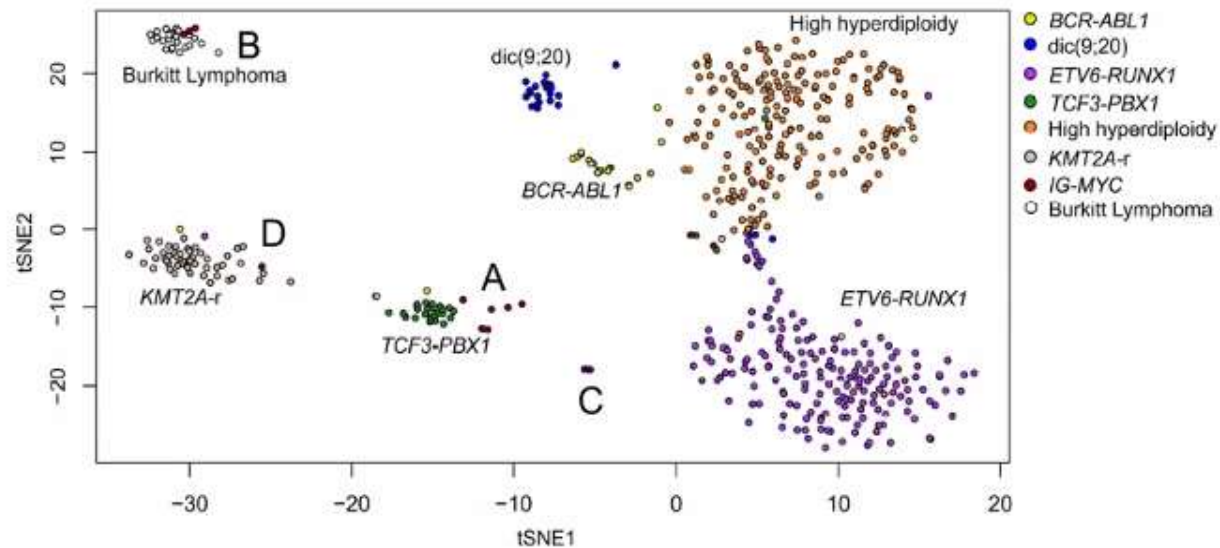
Molecular characterisation and clinical outcome of B-ALL with IG::MYC rearrangement

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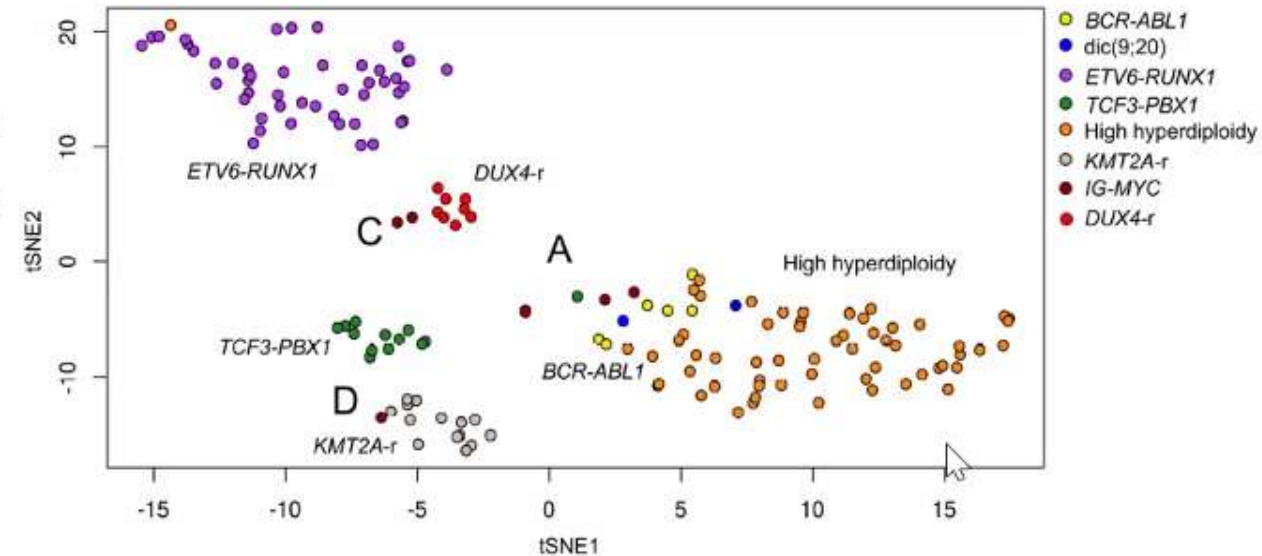
Coexistence with an ALL-specific CA (n=14)

Methylome (5/14) and Transcriptome (6/14) analyses

Methylation data



RNAseq data



NB : 3 cases co-cluster with Burkitt AL/LL

Update of the genetic guidelines – Incorporate *IG::MYC* screening

Patients with HR genetics (KMT2A-r, NH, HoL, iAMP21 or **IG::MYC**) are not eligible for treatment de-intensification and should hence be allocated to **IR-high** group unless their TP1 MRD is >5% in which case they should be allocated to the **HR group**.

-> **IG:: MYC fusions must be confirmed by FISH.**

We strongly recommended using MYC break apart probes such as Cytocell Break apart (LPS027).

On reflection we have decided **not to recommend a specific MYC probe**. *It is out of sync with rest of the guidelines.*

However, we have to keep in a genetic statement about ensuring the chosen probe spans the whole region to detect variable breakpoints.

-> It is **mandatory to exclude rearrangement of BCL2 and/or BCL6 using FISH in all MYC-r**

-> **Identification of MYC- r should be followed by confirmation of the IG partner locus (IGH,IGK,IGL)**

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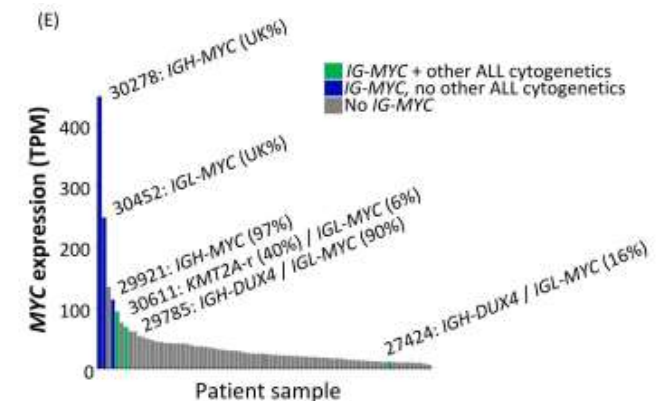
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-> **Identification of MYC- r should be followed by confirmation of the IG partner locus (IGH,IGK,IGL)**

NB :

Ideally, all BCP ALL patients should be screened for *IG::MYC* but a **hierarchical strategy is acceptable** because the stratification is based on the primary CA.

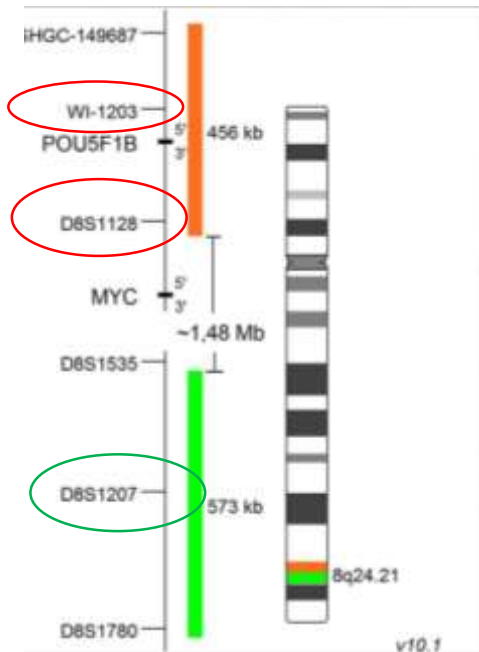
However, the identification of *IG::MYC* **must not prevent screening for other abnormalities**.



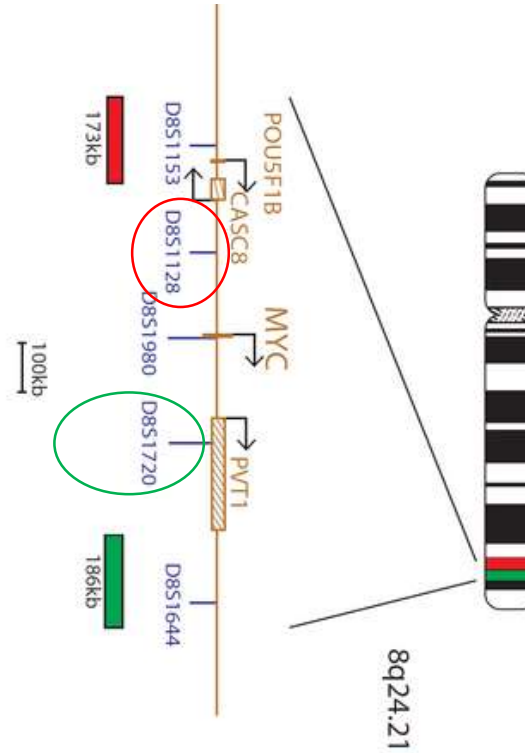
MYC break apart probes

Sonde MYC qui couvre tous les points de cassures (Metasystem, CytoCELL(sysmex), Vysis)

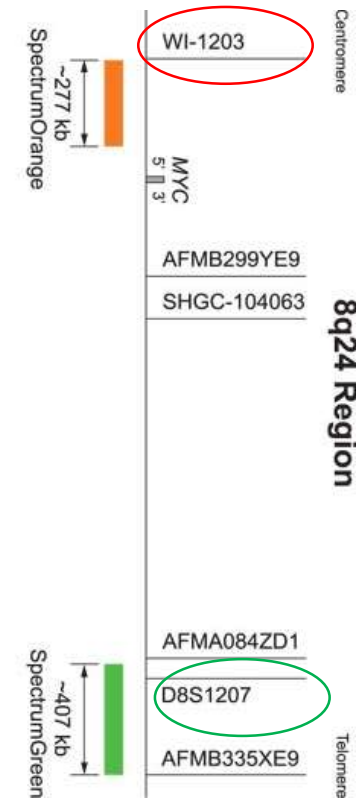
Metasystems



CytoCELL



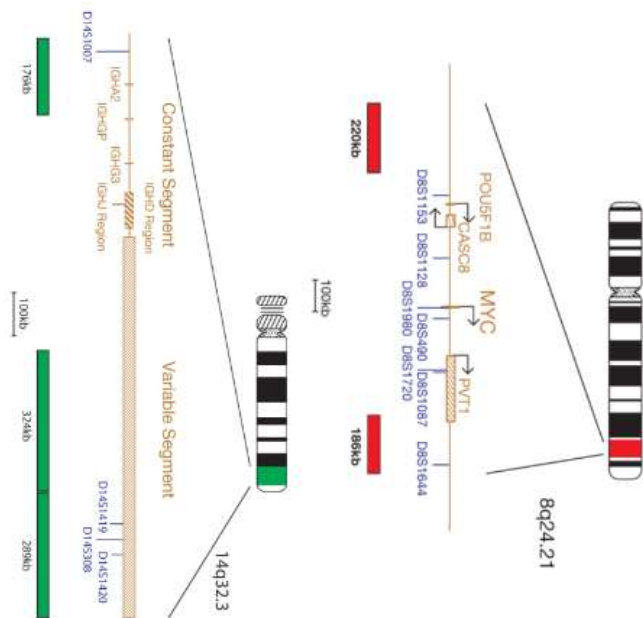
Vysis Abbott



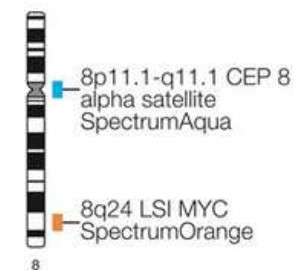
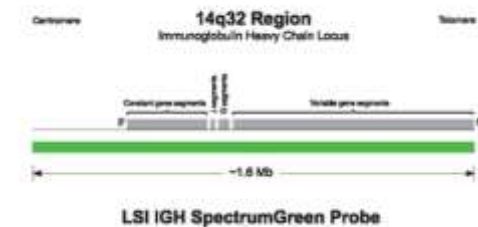
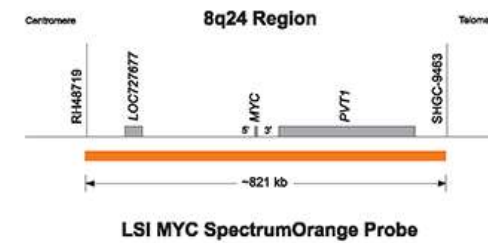
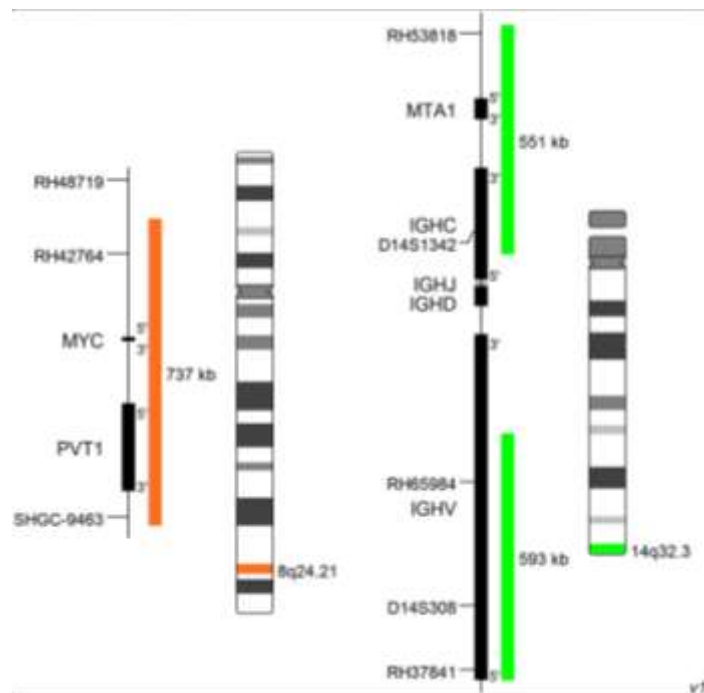
IGH::MYC Dual fusion probes
(CytoCELL, Metasystem, Vysis,)

Vysis Abbott

Cytocell

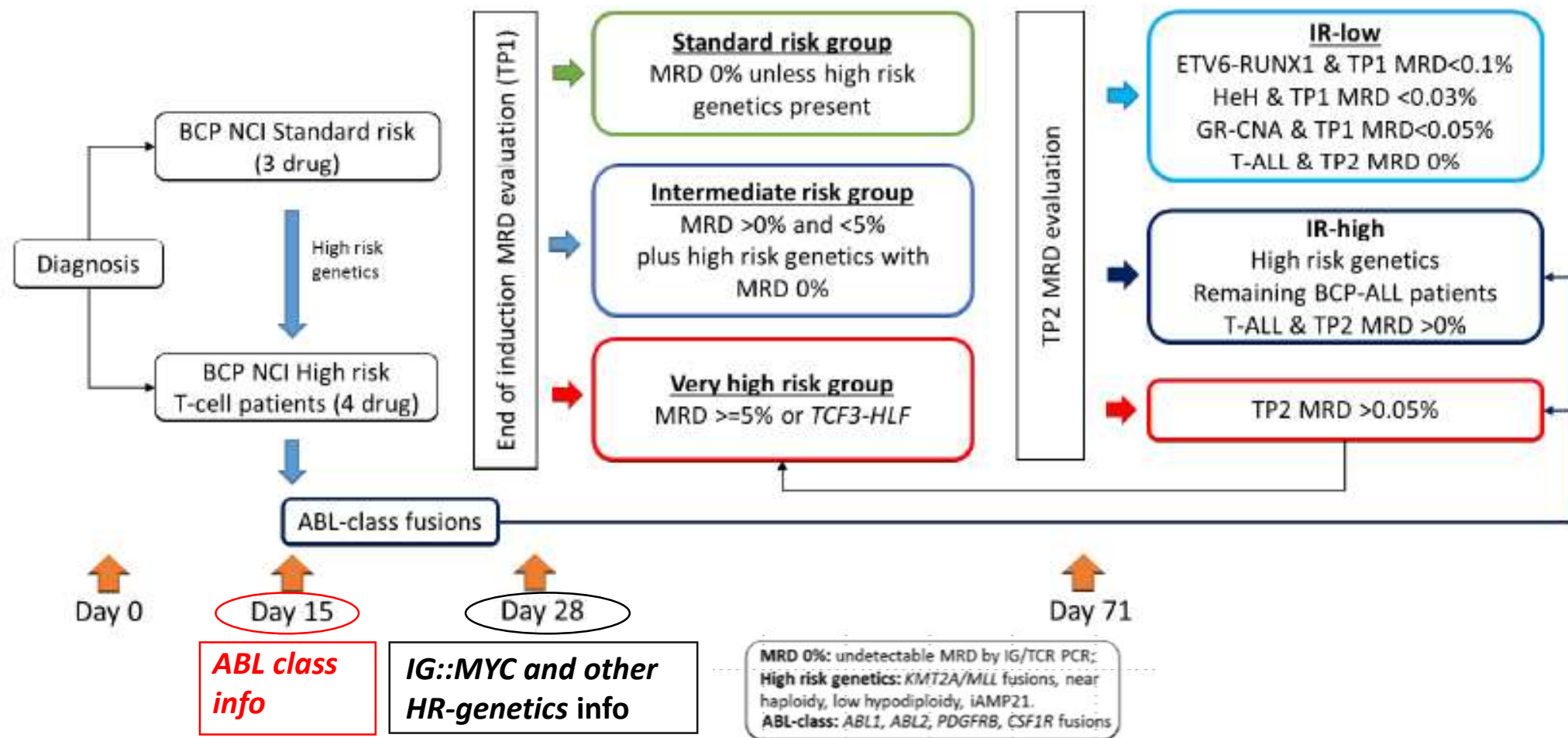


Metasystem



Update of the genetic guidelines – Incorporate *IG::MYC* screening

Patients with HR genetics (*KMT2A*-r, NH, HoL, *iAMP21* or *IG::MYC*) are not eligible for treatment de-intensification and should hence be allocated to **IR-high** group unless their TP1 MRD is >5% in which case they should be allocated to the **HR group**.



ALLTogether **B-ALL**: La FISH en pratique

Panel 1 : type LAL-B «classique» (résultat avant J15)

- ***ETV6::RUNX1***
- ***BCR::ABL1***
- ***KMT2A***
- ***TCF3***

Panel 2, si panel 1 non informatif (résultat pour J15)

- ***ABL-class***
 - *ABL1 (si ABL1 anormal avec BCR::ABL1)*
 - *PDGFRB/CSFR1*
 - *ABL2*
- ***IG::MYC***
 - *MYC ou IGH::MYC*

Panel 3 , si *MYC* pos (résultat pour J28) :

- *BCL2 et BCL6* (si positif : cas exclus du protocole)
- Si *IGH* négatif : *IGK et IGL*

ALLTogether **B-ALL** : La FISH en pratique

Screening des ABL-class

Panel 1 : type LAL-B «classique» (résultat avant J15)

- *ETV6::RUNX1*
- *BCR::ABL1*
- *KMT2A*
- *TCF3*

Panel 2, si panel 1 non informatif (résultat pour J15)

- ***ABL-class ****
 - *ABL1 (si ABL1 anormal avec BCR::ABL1)*
 - *PDGFRB/CSFR1*
 - *ABL2*
- *IG::MYC*
 - *MYC*
 - *IGH::MYC*

Panel 3 , si *MYC* pos (résultat pour J28) :

- *BCL2 et BCL6* (si positif : cas exclus du protocole)
- Si *IGH* négatif : *IGK et IGL*

- *Sauf pour les hypodiploidies mais y compris les Hyperdiploidies élevées*
- *Voir à l'usage, MAIS l'idée est de tenter de demander un amendement pour réduire les « Hyper » à des cas Atypiques +++ :*
 - *nombre modal 51-53*
 - *chromosomes surnuméraires atypiques :*
pas de trisomie 17 ou 18 et présence de trisomie 5 ou 20
(Ensheai A, Lancet Haem, 2021)
 - *delIKZF1*

ALLTogether T-ALL: La FISH en pratique

Patients with T-ALL must be screened for BCR::ABL1, ABL-class and KMT2A fusions (1).

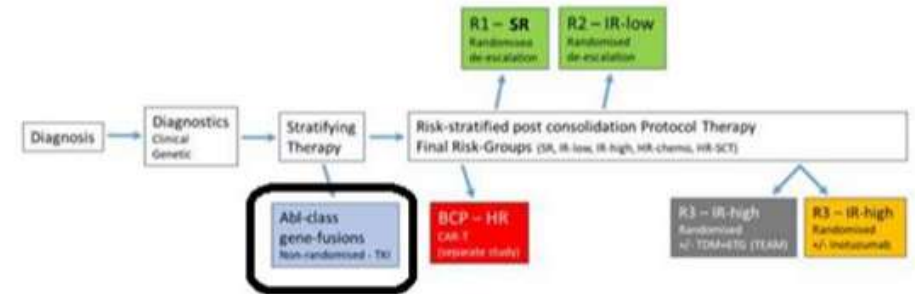
Panel 1 :

- **BCR::ABL1**
- **KMT2A**

Panel 2 : **ABL-class** (resultats pour J15)

- **ABL1** (si ABL1 anormal avec BCR::ABL1)
- **PDGFRB/CSFR1**
- **ABL2**

ALLTogether-1: Overall trial design



At the time of writing these guidelines there is uncertainty as to whether T-ALL with ABL-class fusion should be treated with imatinib.

Hence, we recommend **that all patients with T-ALL are screened for ABL-class fusions** but that treatment with imatinib should be discussed on an individual basis with the national principal investigator. (1)

(1) *Lab guidelines Summary of the recommendations from the ALLTogether Genetics group for the genetic screening of patients treated on the ALLTogether1 Protocol Version 4.0 DDMMM2022*

Table 1: Definition of actionable chromosomal and genomic abnormalities in the ALLTogether paediatric ALL trial

Full name	Abbreviation	Definition	Notes
<i>ETV6::RUNX1</i>	<i>ETV6::RUNX1</i>	<i>t(12;21)(p13;q22)/ETV6::RUNX1</i>	
High hyperdiploidy	HeH	51-67 chromosomes	
<i>KMT2A</i> gene fusions	<i>KMT2A-r</i>	Any genetic rearrangement that results in the fusion of the 5' portion of <i>KMT2A</i> with the 3' portion of a partner gene. The most prevalent partner genes are: <i>AFF1/AF4</i> , <i>MLLT3/AF9</i> , <i>MLLT1/ENL</i> , <i>MLLT10/AF10</i> and <i>MLLT4/AF6/AFDN</i> . Collectively they account for >90% <i>KMT2A-r</i> cases in paediatric ALL.	1) <i>KMT2A</i> was previously named <i>MLL</i> . 2) All <i>KMT2A-r</i> patients will be classified in the same manner; so the identification of the partner gene is not required.
Near-haploidy	NH	Clones with fewer than 30 chromosomes	Doubled-up sub-clones which have 48-58 chromosomes will be included in the subgroup.
Low hypodiploidy	HoL	Clones with 30-39 chromosomes	Doubled-up sub-clones whose karyotypes have 60-78 chromosomes (i.e. near-triploidy) will be included in the subgroup.
Intrachromosomal amplification of chromosome 21	iAMP21	Three or more extra copies of a FISH probe to the <i>RUNX1</i> gene on a single abnormal chromosome 21	iAMP21 can be defined using a DNA array
Primary <i>IG::MYC</i>	<i>IG::MYC</i>	Surface immunoglobulin negative BCP ALL with any of <i>t(8;14)(q24;q32)/IGH::MYC</i> , <i>t(2;8)(p12;q24)/IGK::MYC</i> or <i>t(8;22)(q24;q11)/IGL::MYC</i> demonstrated by FISH	Patients with another actionable chromosomal/genetic abnormality will be stratified according to that other abnormality. Patients with additional rearrangements of <i>BCL2</i> and/or <i>BCL6</i> are excluded from the protocol
<i>t(17;19)(q22;p13)/TCF3::HLF</i>	<i>t(17;19)</i>	<i>t(17;19)(q22;p13)/TCF3::HLF</i>	
Gene fusions involving <i>ABL1</i> , <i>ABL2</i> , <i>PDGFRB</i> and <i>CSF1R</i>	ABL-class fusions	Any genetic rearrangement that results in the fusion of a variable 5' partner gene with the 3' portion of <i>ABL1</i> , <i>ABL2</i> , <i>PDGFRB</i> and <i>CSF1R</i> . Numerous partner genes have been described. The most frequent are <i>EBF1</i> and <i>ETV6</i> .	Patients with <i>BCR::ABL1</i> fusion are excluded from this subgroup and from the protocol. Identification of the partner gene is not required for classification.
Good risk UKALL CNA profile*	CNA-GR	No deletions affecting <i>BTG1</i> , <i>CDKN2A/B</i> , <i>EBF1</i> , <i>ETV6</i> , <i>IKZF1</i> , <i>PAX5</i> [‡] , <i>RB1</i> , and <i>PAR1</i> Isolated deletion of <i>BTG1</i> or <i>ETV6</i> or <i>PAX5</i> [‡] deletion Only two deletions - <i>ETV6</i> and <i>BTG1</i> , <i>ETV6</i> and <i>CDKN2A/B</i> , <i>ETV6</i> and a <i>PAX5</i> [‡] deletion	See Table 4 for details of calling UKALL-CNA profile
Poor risk UKALL CNA profile*	CNA-PR	Any other deletion profile including all deletions of <i>IKZF1</i> , <i>EBF1</i> , <i>RB1</i> and <i>PAR1</i>	See Table 4 for details of calling UKALL-CNA profile

* MLPA was used to develop the CNA profile and is published in Moorman et al (2014) Blood;124(9):1434-44.

‡ For *PAX5*, intragenic amplifications are coded with the deletions as they are predicted to be functionally equivalent.