

ALLTogether

Consortium (ATC) protocol

Cytogenetics STRATIFICATION

GFCH 13 octobre 2022

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GEN-GR

CYTO-GR

or GR-CNA group A

Good risk genetic abnormalities

Good risk cytogenetic abnormalities

- *ETV6-RUNX1/t(12;21)(p13;q22)*
- High Hyperdiploidy (51-65 chromosomes)

Good risk copy number alteration profiles

- No deletion of *IKZF1, CDKN2A/B, PAR1, BTG1, EBF1, PAX5, ETV6 or RB1*
- Isolated deletions of *ETV6, PAX5 or BTG1*
- *ETV6* deletions with a single additional deletion of *BTG1, PAX5 or CDKN2A/B*

GEN-PR

CYTO-HR

or GR-CNA group B

Poor risk genetic abnormalities

High risk cytogenetic subgroups

- *t(9;22)(q34;q11)/BCR-ABL1*
- *MLL/11q23 translocation*
- Near haploidy (<30 chromosomes)
- Low hypodiploidy / near triploidy (30-39 / 60-78 chromosomes)
- Intrachromosomal amplification of chromosome 21 (iAMP21)
- *t(17;19)(q23;p13)/TCF3-HLF*

Intermediate and poor risk copy number alteration profiles

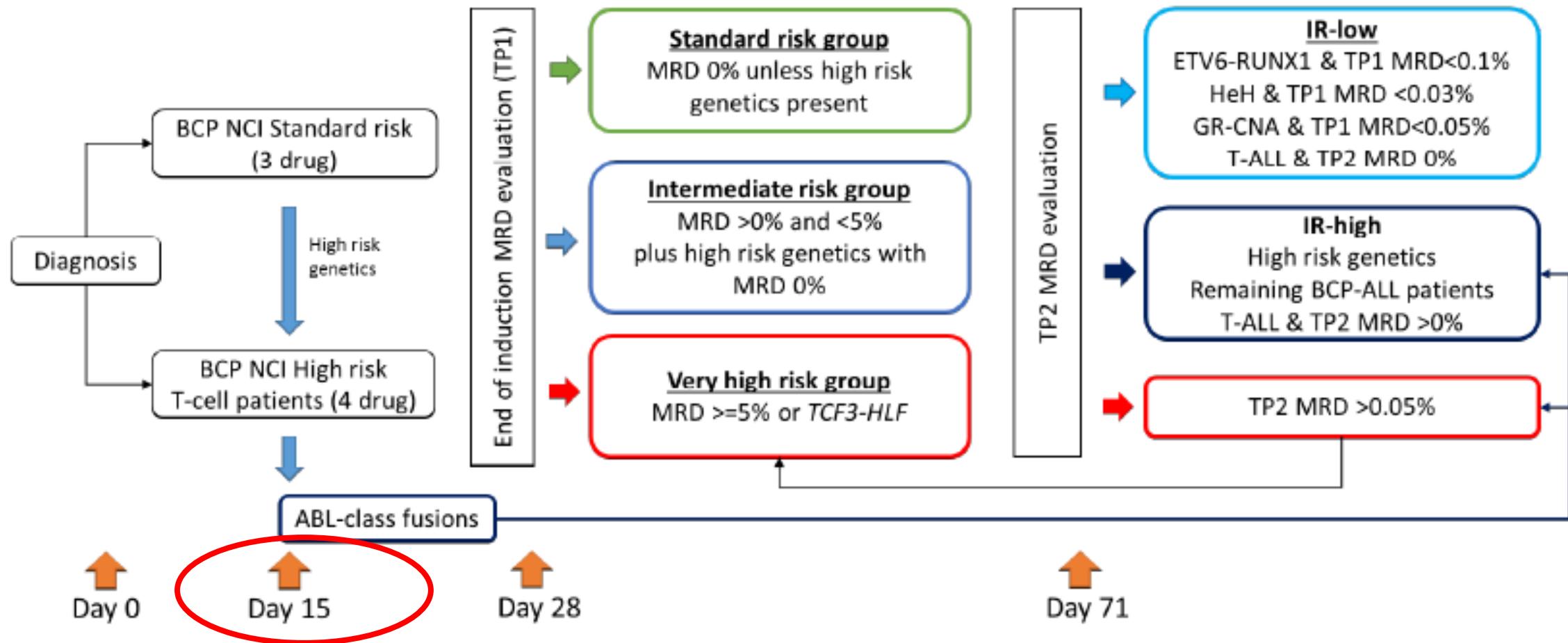
- Any deletion of *IKZF1, PAR1, EBF1 or RB1*
- All other copy number alteration profiles not mentioned above.

Inclusion :
1 à 18 ans

Patients are classified hierarchically with cytogenetic abnormalities taking precedence over copy number alteration profiles.

Definition of novel genetic risk groups for pediatric BCP-ALL.

Moorman AV et al, Blood, 124(9),1434-1444, 2014



MRD 0%: undetectable MRD by IG/TCR PCR;
High risk genetics: *KMT2A/MLL* fusions, near haploidy, low hypodiploidy, iAMP21.
ABL-class: *ABL1*, *ABL2*, *PDGFRB*, *CSF1R* fusions

GR-CNA UKALL profile:

- no deletion of *IKZF1*, *CDKN2A/B*, *PAR1*, *BTG1*, *EBF1*, *PAX5*, *ETV6*, *RB1*;
- isolated deletions of *ETV6*, *PAX5*, *BTG1*;
- ETV6* deletions with a single additional deletion of *BTG1*, *PAX5*, *CDKN2A/B*.

Guidelines for screening methodology in the ALLTogether protocol

Day	Genetic aberrations/karyotypic abnormality patterns	Methods					
		Array	FISH	RT-PCR	MLPA	G/R-Banding	NGS
71	High hyperdiploidy (HeH; 51-67 chromosomes)	X				X	X
15	Low hypodiploidy (HoL; 30-39 chromosomes)	X				X	X
15	Near haploidy (NH; 25-29 chromosomes)	X				X	X
15	iAMP21	X	X ^m				X
15	KMT2A rearrangements		X			X	X
15	BCR-ABL1	X		X		X	X
15	ABL-class fusions (<i>ABL1</i> , <i>ABL2</i> , <i>CSF1R</i> , <i>PDGFRB</i>)	X					X
15	TCF3-HLF	X		X		X	X
71	<i>ETV6-RUNX1</i> , <i>TCF3-PBX1</i> *		X	X		X	X
29/ 71	Copy Number Alterations (Down: <i>IKZF1</i> day 29, rest day 71) <i>BTG1</i> , <i>CDKN2A/B</i> , <i>EBF1</i> , <i>ETV6</i> , <i>IKZF1</i> , <i>PAR1</i> , <i>PAX5</i> , <i>RB1</i>	X			X		X

assumed to be mutually exclusive

* Distinct subgroup, mutually exclusive with other subtypes, with dismal prognosis at relapse

CARYO/FISH pour stratification:

J15

LAL-B

BCR-ABL1 si pas changement de protocole

TEL(ETV6)-AML1(RUNX1) (*iAMP* incluse)

HeH

Low/near hypo

KMT2A

TCF3(E2a) HLF/PBX1

ABL CLASS FUSION : PDGFRB/CSF1R/ABL1/ABL2

(*JAK 2 fait partie des B-others*)

Nouveauté ALLtogether +++

LAL-T

Protocole non modifié avec génétique

BCR-ABL1

KMT2A

Remarque: Anomalie clonale pris en compte théranostic > ou égal à 20% surtout valable pour la biologie moléculaire, mais aussi en Fish....

Point Sondes de FISH:

Première salve :

BCR-ABL1 1^{er} permet check ABL1

TEL-AML1 (+lamp21)

KMT2A(MLL)

E2A(TCF3) 5+/- en fonction centre BM

2eme salve:

3 signaux non trisomie -> **ABL1 break apart**

ABL2 break apart

PDGFRB/CSF1R

Metasystem fait les 2 dans une: gènes collés

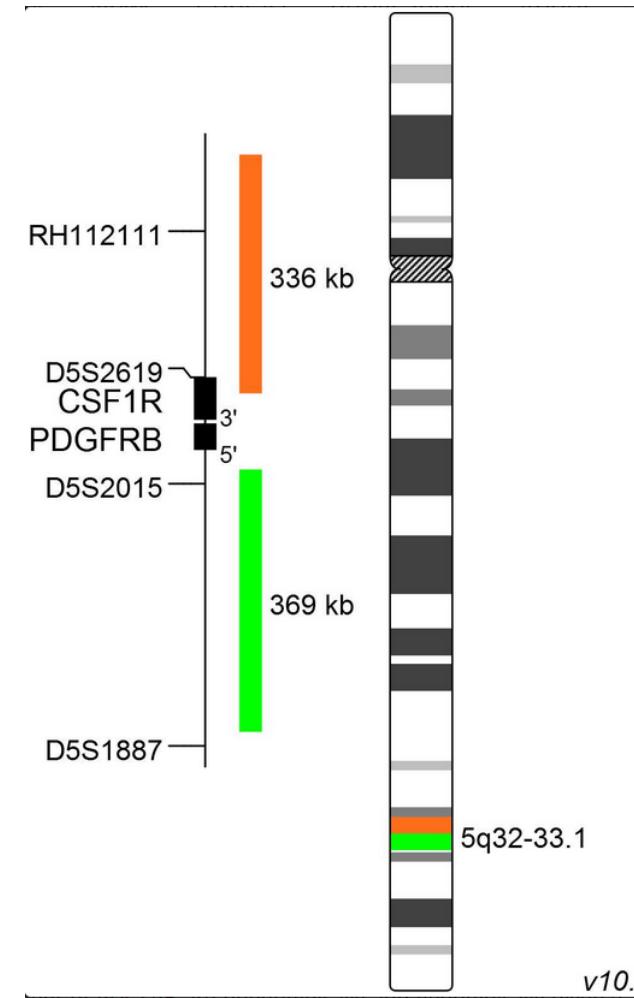
CSFR1 break apart isolée testée (cf infra) utilité?

MAIS PDGFRB/CSF1R suffit à répondre « ABL like » à J15!

JAK2 dans les B other donc post J15

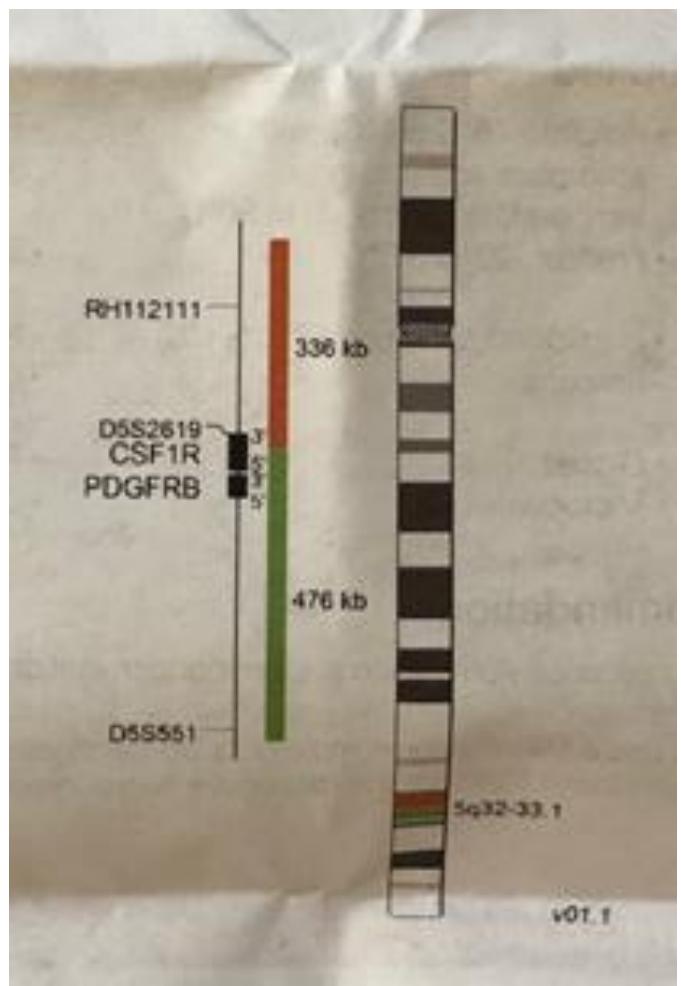
Conclusion:

Tous centres cytogénétiques ayant des LAL-B de l'enfant ALLtogether doit avoir **ABL2 break apart**, et **PDGFRB/CSF1R** (déjà pour SMD/SMP) et **ABL1 break apart** donc en théorie ce sera achat de 2 sondes à prévoir.

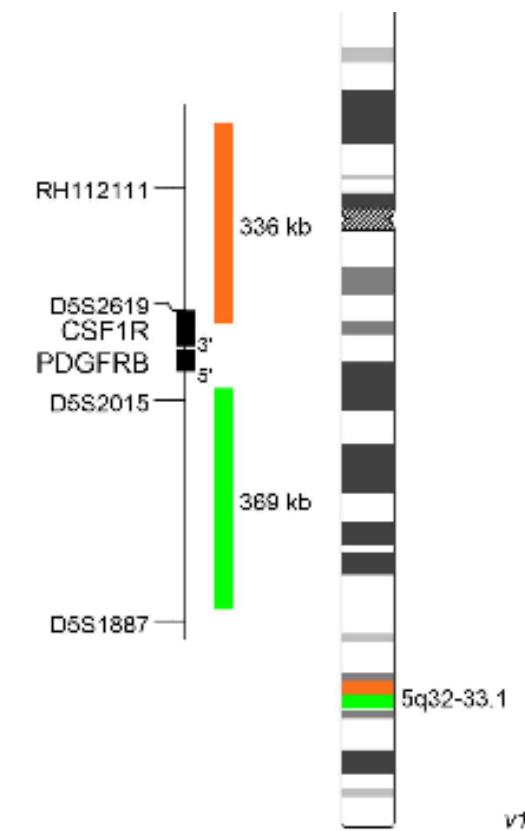


v10.1

CSFR1 Break apart



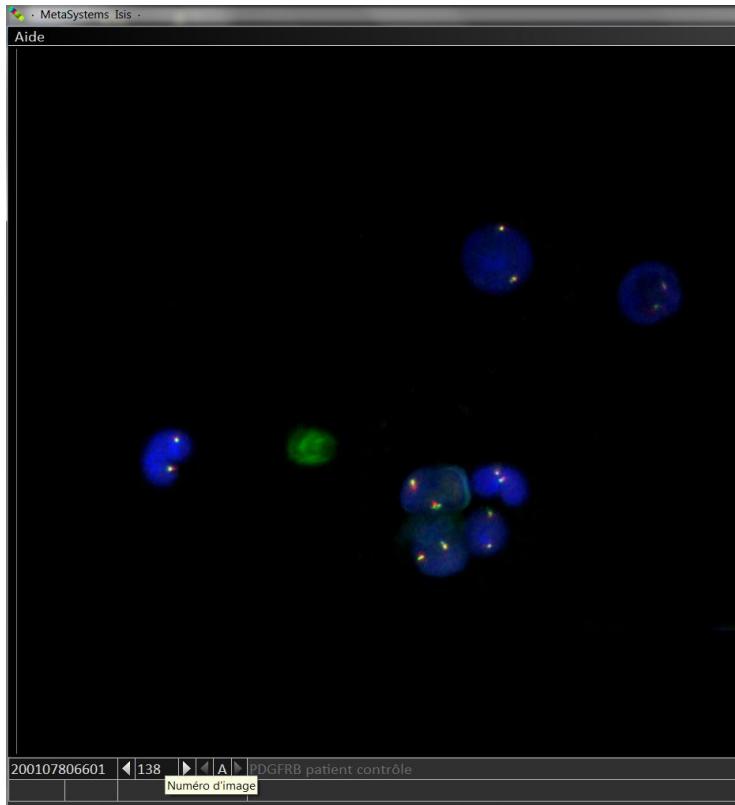
PDGFRB break apart MS



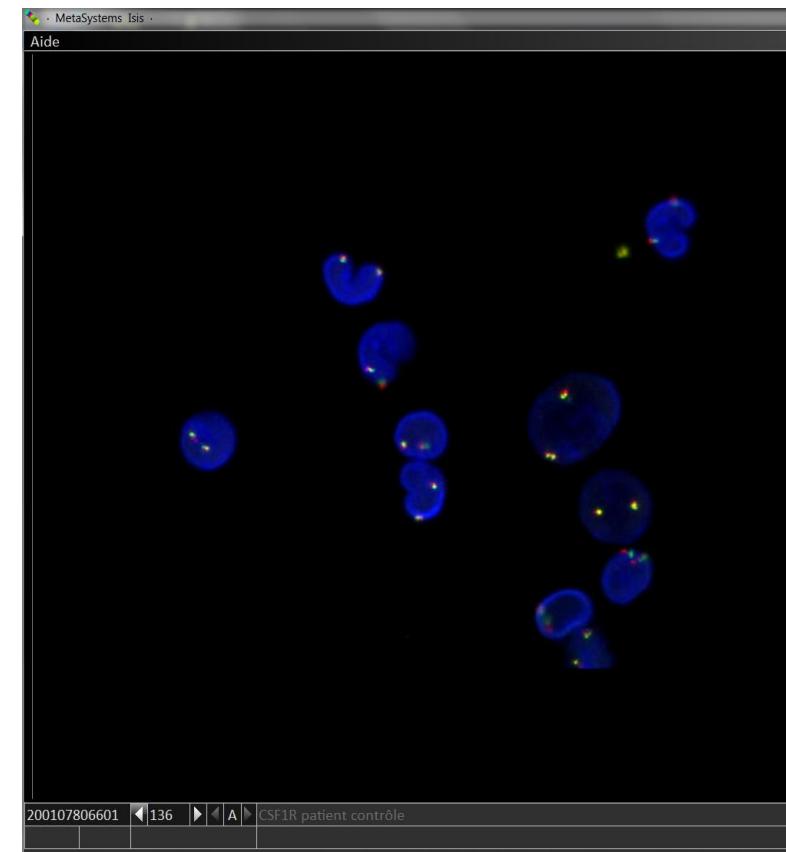
Wendy C

Patient caryotype normal (culot n°987 de 2020)

Absence de réarrangement de PDGFRB/ CSF1R et CSF1R.



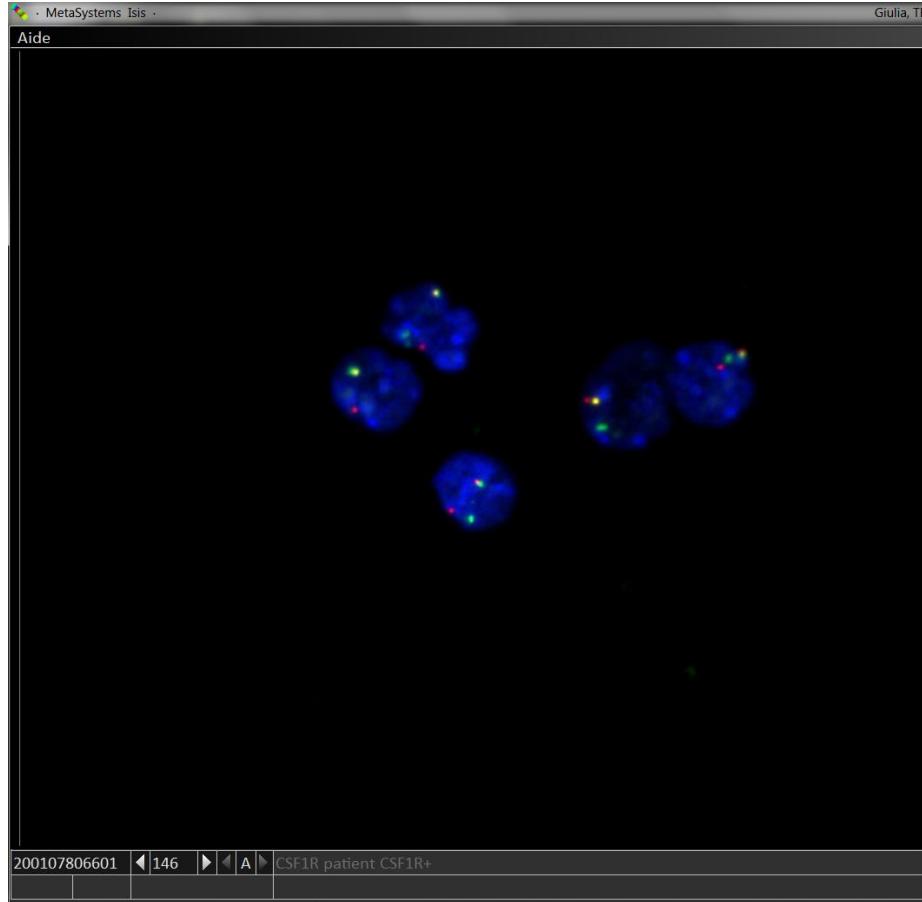
sonde PDGFRB patient contrôle : normal



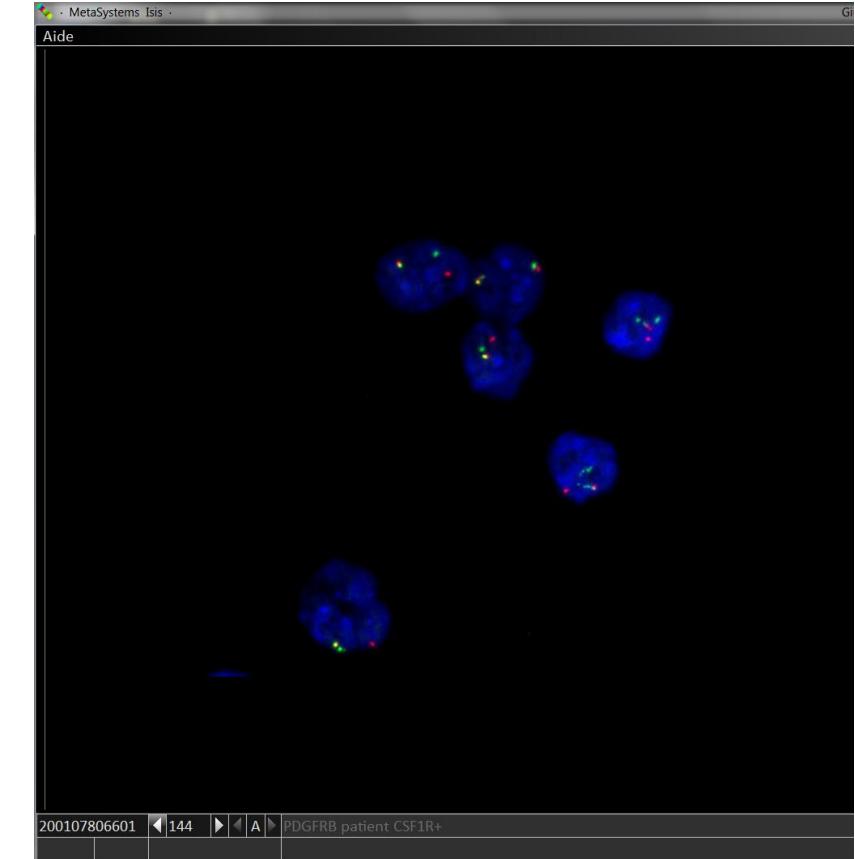
sonde CSF1R patient contrôle : normal

Patient avec réarrangement EBF1-CSF1R en BM

Présence d'un réarrangement avec les 2 sondes PDGFRB/CSF1R et CSF1R BA.



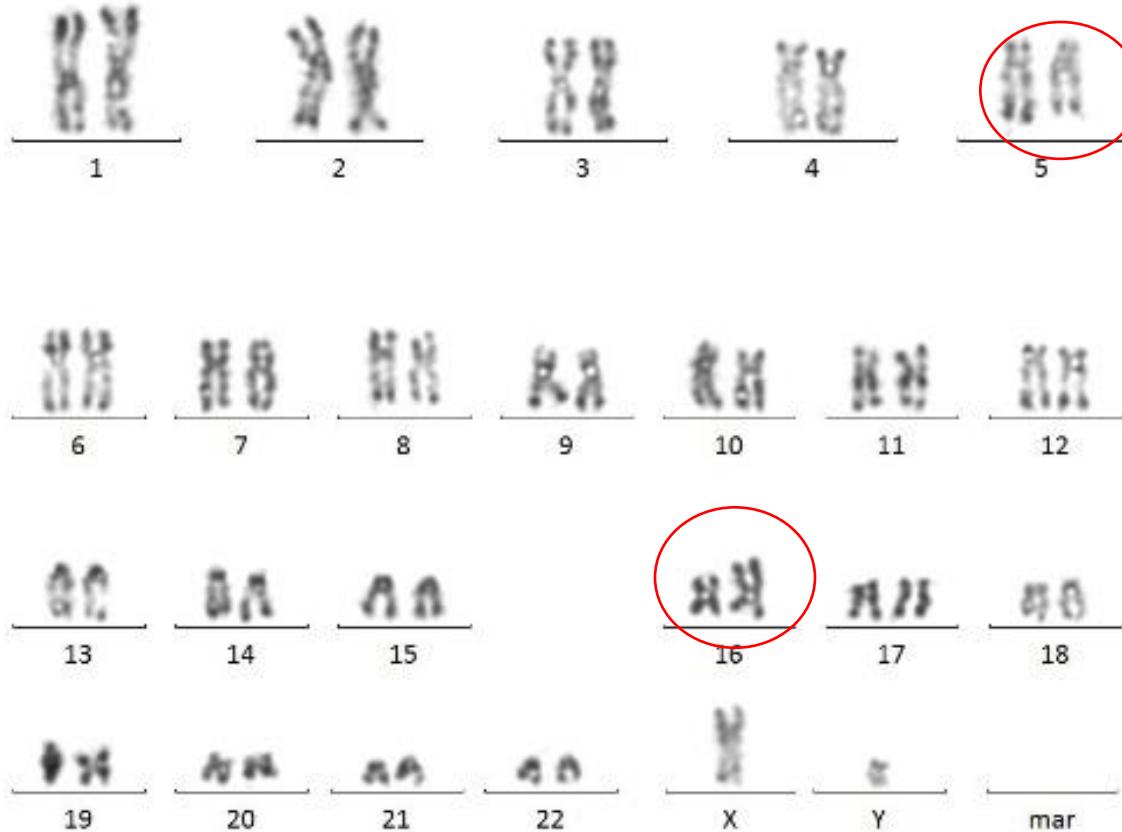
Sonde CSF1 remaniée sur Patient CSF1-EB1 pos



Sonde PDGFRB/CSF1R remaniée sur ce patient CSF1-EB1 pos

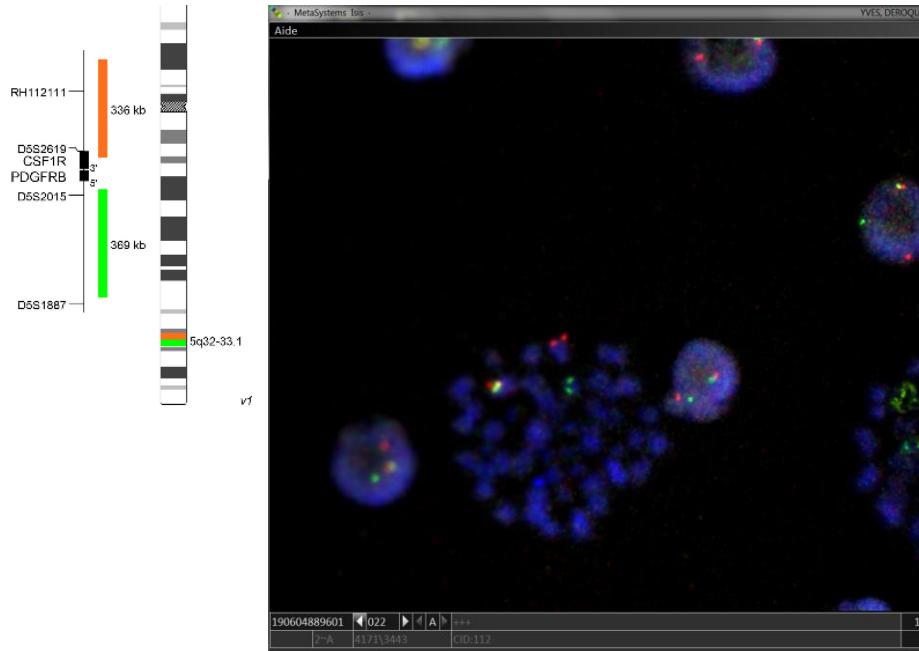
Tester un patient pos PDGFRB transcrit CRAMP1-PDGFRB en BM

Caryotype : 46,XY,t(5;16)(q32;p13)

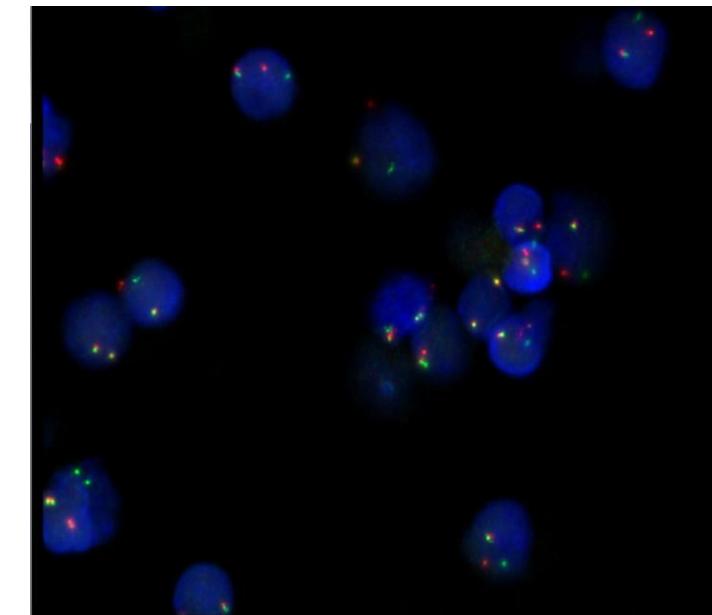
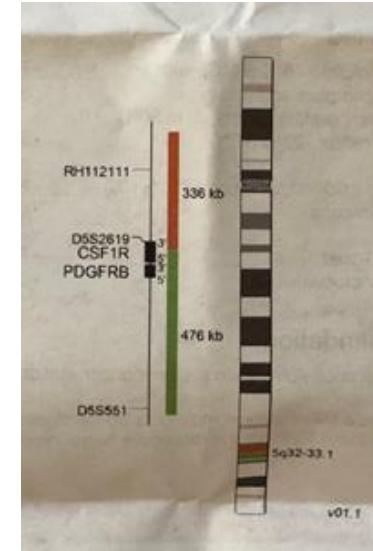


Tester un patient pos PDGFRB transcrit CRAMP1-PDGFRB en BM et caryo t(5;16)

FISH:



Sonde PDGFRB remaniée sur patient PDGFRB CRAMP1 pos avec t(5;16) au caryotype

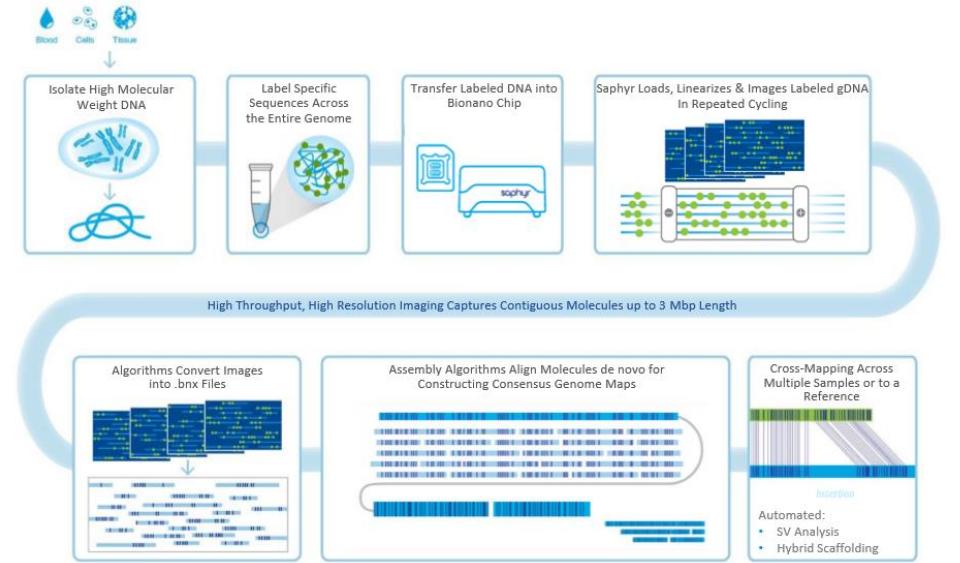


Sonde CSFR1 remaniée sur patient PDGFRB-CRAMP1 MAIS différence dans remaniement indiquant que le dérivé der(5) garde la partie complète centromérique de CSFR1 rouge et petite partie verte (petite fusion), partenaire sur le 16 (partie verte télomérique qui couvre plus PDGFRB donc remaniement de PDGFRB par CSFR1!)

Conclusion : inutile dans protocole ALLtogether le RNAseq donnera le partenaire, on veut juste savoir si « ABL like » la réponse donnée par PDGFRB/CSF1R

Optical genome mapping

- Required viable cells for high molecular weight DNA extraction (>300 kb)
- Fluorescent labelling of DNA molecules used for imaging and mapping to a human genome reference
- Allows genomic aberrations detection with a sensitivity of 5-10% (somatic pipeline):
 - Structural variants : deletion, insertion, inversion, duplication, and translocations (*with a resolution of ~5-10kb*)
 - Copy number variations (CNVs)
 - Aneuploidies
- Previous studies showed a high concordance of OGM results compared to other cytogenetic methods in ALL
- ~10 center within GFCH* testing OGM in hematology
 - (*GFCH Francophone Group of Cytogenetic in Hematology)
- Prospective frontline workflow under current evaluation :
 - Detection of translocations and copy number aberrations required for risk stratification
 - Results expected within 10 days
- If OGM validated : Switch to OGM for cytogenetic results for ALL cases



K Rack, J De Bie, G Ameye, O Gielen, S Demeyer, J Cools, K De Keersmaecker, et al.
« Optimizing the Diagnostic Workflow for Acute Lymphoblastic Leukemia by Optical Genome Mapping ». *American Journal of Hematology* 97, n° 5 (mai 2022): 548-61. <https://doi.org/10.1002/ajh.26487>

V Lestringant, N Duployez , D Penther , I Luquet, C Derrieux, A Lutun, et al. « Optical genome mapping, a promising alternative to gold standard cytogenetic approaches in a series of acute lymphoblastic leukemias ». *Genes Chromosomes Cancer*. oct 2021;60(10):657-67

ACCORD ALLtogether
Suite demande du Pr HELENE CAVE 28/02/22 :
OK OGM si méthode VALIDEE/ACCREDITEE(EEQ)

OGM analysis of an illustrative B-ALL case

